Results: The extent of the oxidative stress and lipid peroxidation is evidenced by significantly higher concentrations of MDA produced by membrane phospholipids in normal tissues adjacent to carcinoma than of normal tissues adjacent to fibroadenoma. Moreover, data obtained from TBA assay on free arachidonic acid (50 μ M) after O.N incubation at 37°C with H₂O₂ (1%) produced more high level of MDA than control. However analysis by ES-MS and MALDI-TOF spectrometry showed the presence of an intermediate radical product of arachidonic acid metabolism only in normal tissues adjacent to breast cancer.

Conclusions: Oxidations of arachidonic acid by reactive oxygen radicals generate a complex family of oxidized lipids known as isoeicosanoids. In our experiments demonstrated that in lipid chloroform extract by normal tissues adjacent to carcinoma presented a anomalous fragmentation pattern of arachidonic acid.

Marine compound

513 POSTER

Pharmacokinetic evaluation of a novel anti-tumor agent, PM01120

J. Yin¹, P. Aviles², M.J. Guillen², C. Ly¹, W. Lee¹, A. Soto², S. Munt², C. Cuevas², G. Faircloth¹. ¹PharmaMar USA, Inc., Cambridge, MA, USA; ²PharmaMar S.A., Colmenar, Madrid, Spain

Introduction: PM01120 is a new synthetic anti-tumor agent related to Variolins that were originally discovered in Antarctic sponge, *Kirkpatrickia variolosa*. PM01120 has demonstrated encouraging results against a panel of human leukemic, ovarian and colon carcinoma cell lines, and multi-drug resistant cell lines. Our recent efforts have focused on the pharmacokinetic evaluation of PM01120 in preclinical species.

Methods: The pharmacokinetic studies were conducted in CD-1 mice, SD rats, New Zealand white rabbits and beagle dogs following a single intravenous or oral dose of PM01120. Blood was collected and plasma sample was harvested for analysis. Brain tissues were also collected from mice PK study to explore the exposure of PM01120 in brain. The plasma samples were processed by liquid-liquid extraction (LLE); the brain tissues were homogenized in buffer and the homogenates were processed by LLE. A liquid chromatography/tandem mass spectrometry assay was used to determine PM01120 concentration in plasma samples or brain tissues. In vitro plasma protein binding of PM01120 was conducted using an ultracentrifugation method. In vitro metabolism study was performed in the liver microsomes from various species.

Results: In mice, PM01120 plasma concentration vs time profile indicated multi-compartmental kinetics after a 5-mg/kg i.v. administration. The mean C_{max} was 1.91 mg/mL and AUC was 0.57 mg hr/mL. The plasma clearance (CL_p) was 147.1 mL/min/kg, which is higher than mouse hepatic blood flow (~90 mL/min/kg). The volume of distribution at steady state (Vdss) was 2.3 L/kg, indicating moderate tissue distribution. The terminal half-life (t_{1/2}) was 6.27 hours. In mice, PM01120 rapidly transferred into brain and peak concentration in brain tissues was reached in less than 5 minutes after i.v. bolus dose. After peak time, brain PM01120 concentrations declined in the similar manner as in circulatory system. The values of C_{max} and AUC were ~1.5-fold higher in brain tissue, compared to PM01120 plasma level. PM01120 showed 39% absolute bioavailability in CD-1 mice after a 10-mg/kg oral administration. In rats, plasma PM01120 concentrations declined with $t_{1/2}$ of 4.32 hours after a 2.5-mg/kg i.v. dose. The mean CL_p was $16.62\,mL/min/kg$ and Vd_{ss} was $0.61\,L/kg$. In rabbits, plasma PM01120 concentrations declined with $t_{1/2}$ of 13-15 hours after a 2.5-mg/kg i.v. dose. The mean CL_p was 22.58-30.04 mL/min/kg and Vd_ss was 1.41-1.42 L/kg. In dogs, plasma PM01120 concentrations declined with $t_{1/2}$ of 0.85-1.05 hours after a 0.5-mg/kg i.v. dose. The mean CL_p was 44.37-44.38 mL/min/kg and Vdss was 1.78 L/kg. The in vitro studies demonstrated that PM01120 was stable in plasma and the plasma protein binding of PM01120 was high in all species studied, ranging from 89 to 98%. In man, the %bound was about 98.91% and this binding was independent of the drug concentration (80 to 800 mM range). In vitro metabolic stability studies showed that PM01120 was metabolized in a moderate to high degree in the presence of NADPH-regenerating system; the in vitro half-lives were 17.93, 6.31, 33.83, 9.90, 9.28, 12.59, and 43.73 min in human, mouse, rat, guinea pig, rabbit, dog, and monkey liver microsomes. The availability of i.v. pharmacokinetic data from four species allowed interspecies scaling of PM01120 to be done, which produced the following relationship between plasma clearance and body weight: $CL_p = 30.506$ (weight^{1.0232}). The extrapolated plasma clearance of 33.67 mL/min/kg was predicted for the average human (70 kg weight). Conclusions: Pharmacokinetic properties were established for PM01120

Conclusions: Pharmacokinetic properties were established for PM01120 in preclinical species. PM01120 showed good oral bioavailability and brain exposure in mice. Interspecies scaling projects a plasma clearance of 33.67 mL/min/kg in humans.

POSTER

Antitumor activity of aplidin® in human neuroblastoma tumors

H. Sasak¹, D. LePage¹, W. Grant¹, M. Elices¹, B. Rinehart¹, T. Caylor¹, L. Cheney¹, K. Halley¹, P. Aviles², G. Faircloth². ¹PharmaMar USA, Pharmacology, Cambridge, MA, USA; ²PharmaMar SA, R&D, Madrid, Spain

Background: Aplidin® (APL), a natural compound originally derived from the Mediterranean tunicate *Aplidium albicans*, is an anti-cancer agent currently in Phase II clinical trials for multiple indications in Europe and the USA. In these trials a suggestion of activity was observed in pediatric neuroblastoma. This observation prompted us to evaluate the cytotoxicity of Aplidin against a broad spectrum of pediatric neuroblastoma human tumor cell lines

Materials and Methods: All cell lines were seeded at two densities in 96 well microtiter plates at 10,000 and 15,000 cells per well. Plates were incubated for 24 hours at 37 degree C prior to treatment for 72 hours with Aplidin® at a concentration range from 10 uM to 26 pM. The degree of cytotoxicity was determined by MTS assay (Tetrazolium), which is based on metabolic reduction of MTS to formazan product that is soluble in the tissue culture medium. The quantity of formazan is measured by the amount of 490 nm absorbance and it is proportional to the number of living cells. The IC $_{50}$, which is an approximate equivalent of IG $_{50}$ (50% growth inhibition) was calculated and converted that to molar concentration by dividing by the molecular weight of Aplidin®.

Results: Aplidin[®] demonstrated significant nanomolar or lower potency in all neuroblastoma cell lines tested (table).

Cell line	IC ₅₀ M/10,000 cells	IC ₅₀ M/15,000 cells
SK-N-AS	2.6 to 1.3 nM	4.4 to 1.1 nM
SK-N-DZ	2.4 nM	4.6 to 1.1 nM
SK-N-MC	13 pM	2.9 to 4.8 nM
SK-N-SH	1.9 to 0.87 nM	2.2 to 1.1 nM

This level of *in vitro* activity was followed up in xenograft studies. Several human neuroblastoma cell lines were implanted subcutaneously into athymic nude mice. Tumors were allowed to grow *in situ* until they reach a size of approximately 100 mm³ at which time animals were randomized into either treatment or control groups.

A detailed analysis of preclinical data with APL will be presented at this venue.

515 POSTER

Transcriptional profiling of palmerolide A, a putative inhibitor of V-ATPase, indicates perturbation of cholesterol biosynthesis

A. Monks¹, N. Reifsnider¹, R. Shoemaker¹, J. Cardellina¹, B. Baker².

¹NCI-Frederick, Screening Technologies Branch, Frederick, USA;

²University of South Florida, Department of Chemistry, Tampa, USA

Palmerolide A (NSC 730282), a 20-membered macrocyclic polyketide bearing carbamate and vinyl amide functionality, was isolated from the tunicate Synoicum adareanum collected from the vicinity of Palmer Station on the Antarctic Peninsula. Palmerolide A was tested in the NCI 60 cell line screen and displayed differential growth inhibition (GI₅₀ ranging from 10 nM to 30 $\mu\text{M})$ and potent toxicity towards several of the melanoma cell lines. The NCI COMPARE algorithm correlated the toxicity profile of palmerolide to several vacuolar ATPase inhibitors, and ongoing studies have demonstrated that palmerolide A inhibits V-ATPase with an IC50 of 2 nM. Two melanoma cell lines (UACC62 and LOX) were treated with 100 nM and 10 nM palmerolide for 6 and 24h, then transcriptional profiles were measured on 35K spot, whole genome, oligonucleotide microarrays (NCI, ATC, Gaithersburg MD), and the data analysed through the mAdb bioinformatics website of the NCI's Center for Cancer Research. Overall, 10 nM (24h) and 100 nM (6h) palmerolide treatment had a limited response on the transcriptome. Gene expression profiles of UACC62 and LOX cell lines after duplicate palmerolide treatments (100 nM) for 24h were well reproduced. A drug-induced change of >1.8-fold in gene expression selected a group of 991 genes which correlated with a Pearson correlation coefficient (PCC) = 0.941 between replicate samples in the UACC62 cell line and PCC=0.628 in the LOX cell line. The LOX cell line was less sensitive to the cytotoxicity of palmerolide and this was borne out by the array analysis where the magnitude and the number of genes dysregulated by palmerolide in LOX was less than in UACC62. Analysis of these 24h treatment data indicated a subset of 169 genes altered >3 fold by 100 nM palmerolide. Functional analysis